

9-Deaza-5'-noraristeromycin

Meral Tuncbilek[†] and Stewart W. Schneller*

Department of Chemistry, Auburn University, Auburn, AL 36849, USA

Received 2 October 2002; accepted 17 March 2003

Abstract—9-Deaza-5'-noraristeromycin (**2**) has been prepared in 10 steps from the readily available (+)-(1*R*,4*S*)-4-*t*-butyldimethylsilyloxycyclopent-2-en-1-yl acetate. Compound **2** was evaluated against a large number of viruses. No activity was found nor did **2** display cytotoxicity towards the viral host cells.

© 2003 Elsevier Science Ltd. All rights reserved.

Introduction

For the past 12 years, we¹ have been investigating the antiviral properties of 5'-norcarbanucleosides of which 5'-noraristeromycin (**1**)² and analogues therefrom represent the most promising series of compounds. Missing from the latter group is the 9-deaza derivative **2**, which is a carbocyclic C-nucleoside.³ In light of the biological properties of 9-deazaadenosine,⁴ we were encouraged to pursue **2**.³ Its preparation and antiviral analysis is reported here (Fig 1).

Chemistry

Adapting a published procedure,⁵ the synthesis of 9-deaza-5'-noraristeromycin began with the reaction of (+)-(1*R*,4*S*)-4-*t*-butyldimethylsilyloxy-2-cyclopenten-1-yl acetate (**3**),⁶ readily available from (+)-(1*R*,4*S*)-4-hydroxy-2-cyclopenten-1-yl acetate (**4**),⁷ with ethyl cyanoacetate in the presence of Pd(O) to give **5**. Oxidation of **5** with osmium tetroxide to **6** was followed by acetonide **7** formation under standard conditions (Scheme 1).

Selective reduction of **7** with diisobutylaluminum hydride gave **8**. Condensation of **8** with aminoacetonitrile bisulfate led to the pyrrole precursor **9**. Protection of the amino nitrogen of **9** as a carbamate with ethyl chloroformate was followed by ring closure with 1,5-diazabicyclo[4.3.0]non-5-ene (DBN) and subsequent

deprotection of the resultant pyrrole nitrogen to avail the substituted pyrrole **10**. Reaction of **10** with formamidine acetate yielded the requisite pyrrolo[3,2-*d*]pyrimidine **11**. Disilylation of **11** to **12** was followed by acid (HCl) deprotection to provide the target **2** as its hydrochloride. Numerous attempts to isolate **2** as the free base failed. X-ray analysis confirmed the structure of **2**.

Antiviral Results

Compound **2** was subjected to antiviral analysis⁸ and found to be inactive. It also showed no cytotoxicity towards the viral host cells.⁹

Conclusion

While 3-deaza-¹⁰ and 7-deaza-5'-noraristeromycin¹¹ displayed antiviral activity, the 9-deaza analogue **2** was ineffective, similar to that reported^{5b} for 9-deazaaristeromycin. This result may be due to the stereoelectronic alteration that accompanied changing from the sp³ N-9 nitrogen of **1** to the sp² carbon of **2**, manifesting

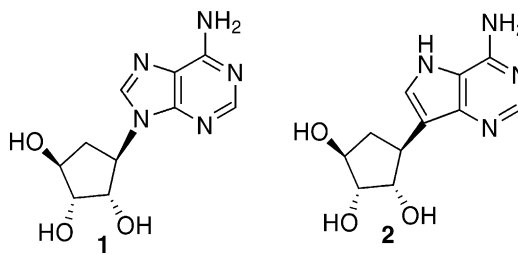
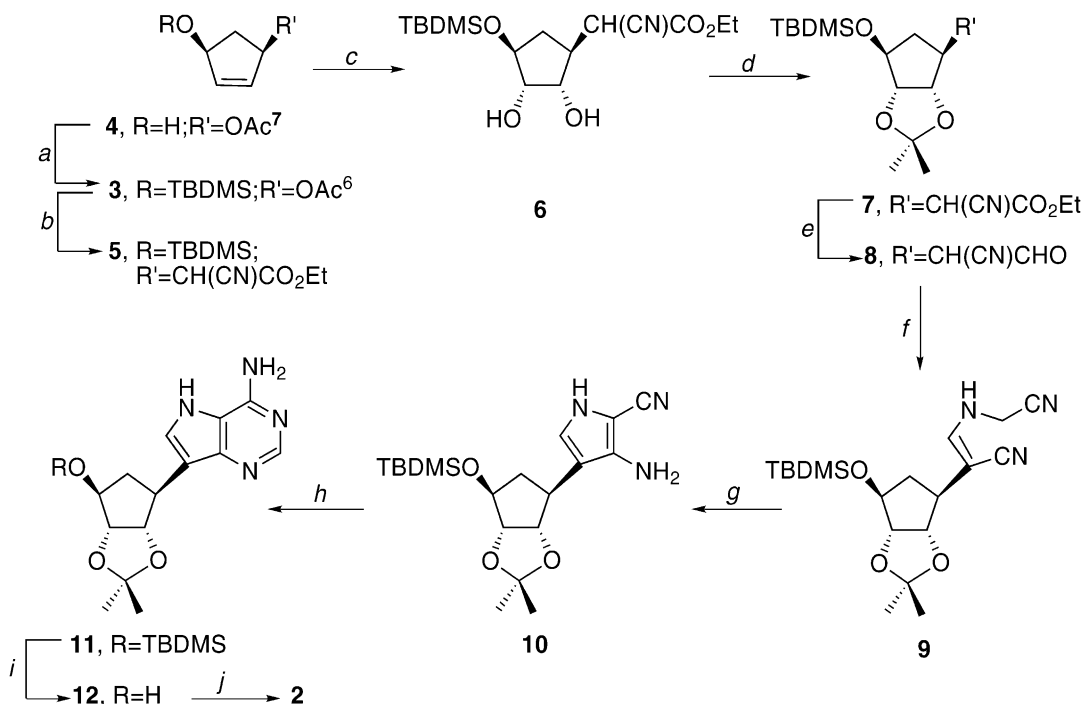


Figure 1.

*Corresponding author. Tel.: +1-334-844-5737; fax: +1-334-844-5748; e-mail: schnest@auburn.edu

[†]On leave from Ankara University, Ankara, Turkey.



Scheme 1. Reaction conditions: (a) TBDMSCl, imidazole, DMF, rt^{6b} (81%); (b) NCCH₂CO₂Et and NaH, (Ph₃P)₄Pd/PPh₃, THF, rt then to 55 °C (78%); (c) using **5**, OsO₄, NMO, acetone–H₂O (49%); (d) Me₂C(OMe)₂, acetone, H⁺ (84%); (e) DIBAL-H, Et₂O, –78 °C; (f) H₂NCH₂CN·H₂SO₄, NaOAc·3H₂O, MeOH, rt (29%); (g) (i) ClCO₂Et, DBN, CH₂Cl₂, rt; (ii) Na₂CO₃, MeOH, rt (16% from 7); (h) HC(=NH)NH₂·AcOH, EtOH, reflux (63%); (i) Bu₄NF, THF, rt (91%); (j) 2N HCl, MeOH, rt, 4 h (51%).

in decreased inhibition of *S*-adenosylhomocysteine hydrolase, the likely cause of the antiviral activity of **1**.¹² This conclusion is supported by the lack of any effect of **2** on vaccinia virus.¹³

Experimental

General

Melting points were recorded on a Meltemp II melting point apparatus and are uncorrected. Combustion analyses were performed by Atlantic Microlab, Inc., Norcross, GA, USA. ¹H and ¹³C NMR spectra were recorded on a Bruker AC 250 spectrometer (operated at 250 and 62.5 MHz, respectively) and are referenced to internal tetramethylsilane (TMS) at 0.0 ppm. The spin multiplicities are indicated by the symbols s (singlet), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), m (multiplet), and br (broad). The optical rotation was measured on a Jasco P-1010 polarimeter. Reactions were monitored by thin-layer chromatography (TLC) using 0.5-mm Whatman Diamond silica gel 60-F₂₅₄ precoated plates with visualization by irradiation with a Mineralight UVGL-25 lamp. Yields refer to chromatographically and spectroscopically (¹H and ¹³C NMR) homogeneous materials.

Ethyl (2*R*/5*S*,1'*R*,4'*S*)-2-[4'-(*t*-butyldimethylsilyloxy)cyclopent-2'-en-1'-yl]-2-cyanoacetate (5**).** To a suspension of NaH (0.8 g, 31.79 mmol, 95%, dry) in anhydrous THF (80 mL) in a 200-mL round-bottomed flask kept under N₂ was added ethyl cyanoacetate (3.38 mL, 31.79 mmol).

The mixture was stirred at rt for 40 min. To this solution was added, successively, *tetrakis*triphenylphosphine palladium (0.90 g, 0.78 mmol), triphenylphosphine (0.62 g, 2.3 mmol) and (+)-(1*R*,4*S*)-4-*t*-butyldimethylsilyloxycyclopent-2-en-1-yl acetate (**3**)^{6b} (4 g, 15.62 mmol) in anhydrous THF (70 mL). The flask was immediately transferred to an oil bath, preheated at 55 °C. The mixture was stirred under N₂ at this temperature for 40 h. Next, the reaction mixture was passed through a SiO₂ plug layered with anhydrous MgSO₄ assisted by purified ether (this step removes suspended materials and some of the catalyst). After concentration of the filtrate under vacuum, the residue was purified by silica gel column chromatography (hexanes–EtOAc, 10:1–8:1) to give **5** (3.77 g, 78%) as a colorless oil. ¹H NMR (CDCl₃): δ 0.07 and 0.08 (2s, 2 × 3H), 0.89 (s, 9H), 1.35 (t, 3H), 1.58 (m, 1H), 2.47 (m, 1H), 3.17 (m, 1H), 3.44 (d, 0.46H, *J* = 8.57 Hz), 3.56 (d, 0.54H, *J* = 7.82 Hz), 4.27 (q, 2H), 4.83 (m, 1H), 5.76–5.93 (m, 2H). ¹³C NMR (CDCl₃) δ –4.9, –4.8, 13.9, 17.9, 25.7, 38.1, 38.4, 42.7, 42.8, 44.1, 44.2, 62.5, 76.2, 76.3, 115.6, 131.4, 137.0, 165.3. Anal. calcd for C₁₆H₂₇NO₃Si: C, 62.10; H, 8.79; N, 4.53. Found: C, 62.06; H, 8.85; N, 4.57.

Ethyl (2*R*/5*S*,1'*R*,2'*S*,3'*S*,4'*S*)-2-[4'-(*t*-butyldimethylsilyloxy)-2',3'-dihydroxycyclopent-1'-yl]-2-cyanoacetate (6**).** To a solution of **5** (3.19 g, 10.3 mmol) and 50% aqueous solution of 4-methylmorpholine *N*-oxide (8 mL, 34 mmol) in a mixture of acetone (50 mL) and H₂O (enough to make the solution clear) was added osmium tetroxide (OsO₄) (100 mg) in one portion. After stirring this solution at rt for 31 h, the solution was washed through a plug of silica gel with EtOAc–MeOH (9:1).

The solvents were removed under reduced pressure. The residue was purified by flash column chromatography eluting with hexanes–EtOAc (1.5:1–1:1) to afford 1.75 g (49%) of **6** as a colorless oil. ^1H NMR (CDCl_3): δ 0.07 and 0.09 (2s, $2 \times 3\text{H}$), 0.89 (s, 9H), 1.33 (t, 3H), 1.49 (m, 1H), 2.07 (m, 1H), 2.32 (m, 1H), 2.52 (m, 1H), 3.05 (d, 1H), 3.33 (d, 1H), 3.65–3.87 (m, 2H), 4.07 (m, 1H), 4.32 (q, 2H). ^{13}C NMR (CDCl_3): δ –4.8, –4.7, 14.1, 18.1, 25.9, 34.6, 35.3, 41.2, 43.2, 43.3, 63.3, 74.3, 74.9, 75.9, 76.0, 78.3, 78.4, 115.7, 115.8, 166.1, 166.3. Anal. calcd for $\text{C}_{16}\text{H}_{29}\text{NO}_3\text{Si} \cdot 0.5 \text{ EtOAc}$: C, 55.79; H, 8.50; N, 3.69. Found: C, 55.39; H, 8.58; N, 3.61.

Ethyl (2*R*,1*R*,2*S*,3*S*,4*S*)-2-[4'-(*t*-butyldimethylsilyloxy)-2',3'-isopropylidenedioxycyclopent-1'-yl]-2-cyanoacetate (7**).** To a solution of **6** (2.73 g, 7.9 mmol) in dry acetone (50 mL) and 2,3-dimethoxypropane (6 mL) in a 100 mL round-bottomed flask was added a small crystal of *p*-toluenesulfonic acid under N_2 . This mixture was stirred at rt for 20 h and then was passed through a plug of silica gel assisted by reagent grade acetone. The resulting solution was evaporated under reduced pressure and the residue was chromatographed (hexanes–EtOAc, 6:1) to afford 2.55 g (84%) of **7** as a colorless oil. ^1H NMR (CDCl_3): δ 0.10 and 0.12 (2s, $2 \times 3\text{H}$), 0.90 (s, 9H), 1.26–1.42 (m, 10H), 2.23 (m, 1H), 2.65 (m, 1H), 3.76 (d, 0.62H, $J=11.38\text{ Hz}$), 3.91 (d, 0.38H, $J=11.60\text{ Hz}$), 4.20 (m, 1H), 4.29 (q, 2H), 4.34 (t, 1H), 4.52–4.77 (dd, 1H). ^{13}C NMR (CDCl_3): δ –5.5, –5.4, 13.5, 17.5, 23.6, 25.3, 25.9, 34.1, 34.6, 39.5, 40.5, 45.2, 45.6, 62.3, 62.4, 76.2, 76.7, 82.5, 82.8, 86.4, 110.3, 110.4, 115.6, 115.7, 164.9, 165.2. Anal. calcd for $\text{C}_{19}\text{H}_{33}\text{NO}_5\text{Si}$: C, 59.60; H, 8.67; N, 3.65. Found: C, 59.71; H, 8.74; N, 3.47.

(1*R*,2*S*,3*S*,4*S*)-3-Amino-2-cyano-4-[(4'-(*t*-butyldimethylsilyloxy)-2',3'-isopropylidenedioxycyclopent-1'-yl)-1*H*-pyrrole (10**).** To a solution of **7** (4.01 g, 10.45 mmol) in anhydrous ethyl ether (60 mL) at -78°C was added DIBAL-H (1 M in hexanes, 21 mL) over 15 min. The mixture was stirred at -78°C for 30 min and MeOH (90 mL) was added. The mixture was stirred for 30 min and concentrated. The resultant solid cake was suspended in EtOAc–MeOH (25:2) then stirred for 40 min and filtered. After removal of the filtrate under reduced pressure, the residue containing **8** was dissolved in MeOH (70 mL). Aminoacetonitrile bisulfate (3.98 g, 25.82 mmol) and $\text{NaOAc} \cdot 3\text{H}_2\text{O}$ (3.39 g, 24.9 mmol) were added. The mixture was stirred at rt for 43 h. The solvent was evaporated at rt and the residue was co-evaporated with MeOH ($3 \times 60\text{ mL}$). Ethyl acetate (200 mL) was added to the residue and this mixture stirred for 45 min. The mixture was filtered, the filtrate concentrated and the residue chromatographed (hexanes–EtOAc, 2:1) to give 1.13 g (29%) of **9** as a white solid. To a solution of **9** (1.13 g, 2.99 mmol) in dry CH_2Cl_2 (55 mL) at 0°C was added DBN (6.46 mL, 52.29 mmol) and ethyl chloroformate (2.99 mL, 31.27 mmol). The mixture was stirred for 1 h at rt and then washed with H_2O . The organic phase was dried (MgSO_4), concentrated and the residue chromatographed (hexanes–EtOAc, 2:1) to give, after evaporation of the solvent, a material that was dissolved in

MeOH (35 mL). Sodium carbonate (40 mg, 0.37 mmol) was added to this solution. The new mixture was stirred 1 h at rt and solvent was removed under reduced pressure. The resultant residue was purified by silica gel column chromatography (hexanes–EtOAc, 4:1) to give **10** (630 mg, 16% from **7**) as a semi-white solid: mp 50°C . ^1H NMR (CDCl_3): δ 0.08 and 0.09 (2s, $2 \times 3\text{H}$), 0.90 (s, 9H), 1.29 (s, 3H), 1.54 (s, 3H), 1.85 (m, 1H), 2.39 (m, 1H), 2.90 (m, 1H), 4.07 (s, 2H), 4.27–4.41 (m, 3H), 6.48 (s, 1H), 7.9 (s, 1H). ^{13}C NMR (CDCl_3): δ –4.6, 18.3, 25.2, 26.0, 27.5, 38.5, 40.4, 85.8, 86.6, 88.3, 112.8, 114.6, 115.2, 120.6, 142.8. Anal. calcd for $\text{C}_{19}\text{H}_{31}\text{N}_3\text{O}_3\text{Si} \cdot 0.3 \text{ EtOAc}$: C, 60.06; H, 8.33; N, 10.40. Found: C, 60.11; H, 8.45; N, 10.13.

(1*R*,2*S*,3*S*,4*S*)-4-Amino-7-[4'-(*t*-butyldimethylsilyloxy)-2',3'-isopropylidenedioxycyclopent-1'-yl]-5*H*-pyrrolo[3,2-*d*]pyrimidine (11**).** To a solution of **10** (610 mg, 1.61 mmol) in EtOH (30 mL) was added formamidine acetate (667 mg, 6.41 mmol) and this mixture refluxed for 8 h. The solvent was then removed by evaporation under reduced pressure and the residue purified via column chromatography, eluting with EtOAc. Fractions containing the product were combined and evaporated to afford 410 mg (63%) of **11** as a white solid: mp 134°C dec. ^1H NMR ($\text{DMSO}-d_6$): δ –0.02 and 0.06 (2s, $2 \times 3\text{H}$), 0.82 (s, 9H), 1.21 (s, 3H), 1.41 (s, 3H), 2.10 (m, 1H), 2.30 (m, 1H), 4.16 (t, 1H), 4.42 (t, 1H), 4.87 (q, 1H), 6.62 (s, 2H), 7.36 (s, 1H), 8.08 (s, 1H), 10.70 (s, 1H). ^{13}C NMR ($\text{DMSO}-d_6$): δ –4.8, 17.7, 21.0, 24.8, 25.6, 27.1, 78.1, 84.2, 87.2, 111.1, 113.9, 115.4, 125.1, 145.1, 149.6, 150.3. Anal. calcd for $\text{C}_{20}\text{H}_{32}\text{N}_4\text{O}_3\text{Si} \cdot 0.6 \text{ CH}_3\text{OH}$: C, 58.33; H, 8.11; N, 13.21. Found: C, 58.10; H, 7.99; N, 12.82.

(1*R*,2*S*,3*R*,4*S*)-4-Amino-7-(4'-hydroxy-2',3'-isopropylidenedioxycyclopent-1'-yl)-5*H*-pyrrolo[3,2-*d*]pyrimidine (12**).** A solution of **11** (400 mg, 0.99 mmol) in anhydrous THF (40 mL) and Bu_4NF (1.0 M solution in THF, 2.8 mL) was stirred at rt for 23 h. The solvent was evaporated under reduced pressure and the residue purified via column chromatography, eluting with EtOAc–MeOH (4:1) to afford 260 mg (91%) of **12** as a white solid: mp 248°C . ^1H NMR ($\text{DMSO}-d_6$): δ 1.20 (s, 3H), 1.41 (s, 3H), 1.99 (m, 1H), 2.37 (m, 1H), 3.23 (m, 1H), 4.04 (m, 1H), 4.41 (q, 1H), 4.78 (q, 1H), 5.67 (br s, 1H), 6.70 (s, 2H), 7.40 (s, 1H), 8.08 (s, 1H), 10.70 (s, 1H). ^{13}C NMR ($\text{DMSO}-d_6$): δ 24.6, 27.1, 75.9, 85.1, 87.2, 110.5, 113.8, 115.8, 125.3, 144.6, 149.4, 150.3. Anal. calcd for $\text{C}_{14}\text{H}_{18}\text{N}_4\text{O}_3 \cdot 0.5\text{H}_2\text{O}$: C, 56.18; H, 6.40; N, 18.72. Found: C, 55.94; H, 6.39; N, 18.53.

(1*R*,2*S*,3*R*,4*S*)-4-Amino-7-(2',3',4'-trihydroxycyclopent-1'-yl)-5*H*-pyrrolo[3,2-*d*]pyrimidine (9-deaza-5'-nor-aristeromycin) (2**) hydrochloride.** Compound **12** (170 mg, 0.58 mmol) was suspended in HCl (2 N solution in MeOH, 35 mL) and this mixture was stirred for 4 h. The solvent was evaporated under reduced pressure and the residue was co-evaporated with MeOH ($3 \times 30\text{ mL}$). This material was recrystallized from MeOH to provide **2** (85 mg, 51%) as a white solid: mp 246°C ; $[\alpha]_{\text{D}}^{23.6} -39.52$ (c 0.084, H_2O). ^1H NMR ($\text{DMSO}-d_6$): δ 1.47 (m, 1H, H-5'), 2.47 (m, 1H, H-5'), 3.20 (m, 1H,

H-1'), 3.71 (m, 1H, H-4'), 3.94 (m, 2H, H-2' and H-3'), 4.4–5.1 (br s, 2H, 2 × OH), 7.74 (d, 1H, H-6), 8.51 (s, 1H, H-2), 8.93 (br s, 2H, NH₂), 12.67 (s, 1H, NH). ¹³C NMR (DMSO-*d*₆) δ 36.8, 37.4, 75.2, 77.6, 77.9, 112.6, 114.3, 128.6, 133.0, 144.7, 152.4. Anal. calcd for C₁₁H₁₄N₄O₃·HCl: C, 46.08; H, 5.27; N, 19.54. Found: C, 46.27; H, 5.31; N, 19.31.

Antiviral assays. The antiviral and toxicity analyses were performed following standard procedures reported by us previously.^{2,14}

Acknowledgements

This research was supported by funds from the Department of Health and Human Services (AI 48495), which is appreciated. We are also indebted to the following individuals for providing the antiviral data^{8,9} following their standard procedures:^{2,14,15} Dr. Erik De Clercq, the Rega Institute, Leuven, Belgium; Dr. Earl Kern, University of Alabama at Birmingham, Birmingham, AL; Dr. Brent Korba, Georgetown University, Washington, DC; and, Dr. Robert Sidwell, Utah State University, Logan, UT. We are grateful to Dr. Thomas Albrecht-Schmitt, Auburn University, for performing the X-ray analysis of **2**.

References and Notes

1. Koga, M.; Schneller, S. W. *Tetrahedron Lett.* **1990**, *31*, 5861.
2. For a leading reference, see Rajappan, V. P.; Schneller, S. W.; Williams, S. L.; Kern, E. R. *Bioorg. Med. Chem.* **2002**, *10*, 883.
3. (a) Boyer, S. J.; Leahy, J. W. *J. Org. Chem.* **1997**, *62*, 3976. (b) Mohar, B.; Kobe, J. *Nucleosides Nucleotides* **1999**, *18*, 443. (c) Kuang, R.; Ganguly, A. K.; Chan, T.-M.; Pramanik, B. N.; Blythin, D. J.; McPhail, A. T.; Saksena, A. K. *Tetrahedron Lett.* **2000**, *41*, 9575.
4. (a) Zimmerman, T. P.; Deeprose, R. D.; Wolberg, G.; Stopford, C. R.; Duncan, G. S.; Miller, W. H.; Miller, R. L.; Lim, M. I.; Ren, W. Y.; Klein, R. S. *Biochem. Pharmacol.* **1983**, *32*, 1211. (b) Namikoshi, M.; Carmichael, W. W.; Sakai, R.; Jares-Erijman, W. W.; Kaup, A. M.; Rinehart, K. L. *J. Am. Chem. Soc.* **1993**, *115*, 2504.
5. (a) Chun, B. K.; Chu, C. K. *Tetrahedron Lett.* **1999**, *40*, 3309. (b) Chun, B. K.; Song, G. Y.; Chu, C. K. *J. Org. Chem.* **2001**, *66*, 4852.
6. (a) Paquette, L. A.; Heidelbaugh, T. M. *Org. Synth.* **1995**, *73*, 44. (b) Basra, S. K.; Drew, M. G. B.; Mann, J.; Kane, P. D. *J. Chem. Soc., Perkin Trans. 1* **2000**, 3592.
7. Siddiqi, S. M.; Chen, X.; Schneller, S. W. *Nucleosides Nucleotides* **1993**, *12*, 267.
8. Viruses subjected to **2** were influenza A (H1N1 and H3N2), influenza B, parainfluenza-3 virus, respiratory syncytial virus, vesicular stomatitis virus, coxsackie virus B4, reovirus-1, herpes simplex virus 1 (TK⁺ and TK⁻) and 2, human cytomegalovirus, Epstein–Barr virus, varicella zoster virus, vaccinia virus, cowpox virus, West Nile virus, Sindbis virus, adenovirus type 1, measles, punta toro virus, rhinovirus type 2, Venezuelan equine encephalitis, yellow fever, and human immunodeficiency virus 1 and 2.
9. Host cells used in the assays: HEL, HeLa, Vero, HFF, CEM, A-549, CV-1, KB, MA-104, and MDCK.
10. Siddiqi, S. M.; Chen, X.; Rao, J.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. *J. Med. Chem.* **1995**, *38*, 1035.
11. Seley, K. L.; Schneller, S. W.; Rattendi, D.; Bacchi, C. J. *J. Med. Chem.* **1997**, *40*, 622.
12. De Clercq, E. *Nucleosides Nucleotides* **1998**, *17*, 625.
13. Cools, M.; De Clercq, E. *Biochem. Pharmacol.* **1989**, *38*, 1061.
14. (a) Siddiqi, S. M.; Chen, X.; Schneller, S. W.; Ikeda, S.; Snoeck, R.; Andrei, G.; Balzarini, J.; De Clercq, E. *J. Med. Chem.* **1994**, *37*, 551. (b) Seley, K. L.; Schneller, S. W.; Korba, B. *Nucleosides Nucleotides* **1997**, *16*, 2095.
15. Barnard, D. L.; Stowell, V. D.; Seley, K. L.; Hegde, V. R.; Das, S. R.; Rajappan, V. P.; Schneller, S. W.; Smee, D. F.; Sidwell, R. W. *Antiviral Chem. Chemother.* **2001**, *12*, 241.